Designing protein-protein interactions with self-supervised geometric deep learning

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Protein—protein interactions



Video source: YouTube channel Vaccine Makers Project

Directly linked to the development and treatment of viruses, stroke, cancer, Alzheimer, ...



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Staphylokinase: thrombolytic drug candidate

Brain image source: Bel Marra Health website





Fibrin clot

Stroke



Fibrin degradation product



Staphylokinase: thrombolytic drug candidate



Brain image source: Bel Marra Health website

Fibrin degradation product





Staphylokinase: thrombolytic drug candidate



How to enhance the *binding affinity* of the interaction for effective thrombolysis?

Brain image source: Bel Marra Health website









What is a protein? Protein: a folded chain of amino acids 20 amino acids (building block types)





Staphylokinase—microplasmin interaction



How to enhance the binding affinity of the interaction?

Staphylokinase—microplasmin interaction



What amino acids of staphylokinase to mutate and how? 20ⁿ combinations

Standard approach: $\Delta\Delta G$ screening

- 1. Screen thousands or millions of mutations according to $\Delta\Delta G$ binding energy change upon mutation ranging roughly in [-12, 12]
- 2. Select several best candidates (with lowest $\Delta\Delta G$) and test in a lab





State of the art for predicting $\Delta\Delta G$



- Often require **mutant 3D structure** → slow
- Weak evaluation protocol → poor generalization

• Rely on small data (7K annotated mutations from SKEMPI2) \rightarrow unstable, weak generalization



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Labeled data (SKEMPI2) 300 interactions, 7K mutations



Protein C, amino acid 21 $Cys \rightarrow Val$ $\Delta\Delta G = -0.025$



Protein I, amino acid 45 Leu \rightarrow Ser $\Delta\Delta G = 1.17$



PPIRef: New large dataset of PPIs

Labeled data (SKEMPI2) 300 interactions, 7K mutations



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Unlabeled data (Protein Data Bank) 322K interactions in our PPIRef, 41K in DIPS







iDist: Scalable comparison of PPIs

- **iDist** accurately approximates iAlign¹⁸ (TM-score for PPIs) (near-duplicate detection with 99% precision and 97% recall)
- **iDist** is ~500 times faster than iAlign

3P9R



iDist = 0.0035

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- Available datasets are redundant and incomplete (Some PPIs represented > 500 times, many missing)
- Existing train-test splits suffer from data leakage (>53% of test sets have near-duplicates in train sets)





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• **PPIRef**: non-redundant complete set of PPIs from PDB



iDist = 0.0035

Input PPI c









PPIformer



→ Self-supervised pre-training → Downstream task fine-tuning

 \rightarrow Both



PPIformer



Self-supervised pre-training — Downstream task fine-tuning

- Leverages big data (millions of masked examples from PPIRef during pre-training)
- Very fast, requires a single forward pass on the native 3D structure
- Fine-tuned and evaluated on non-leaking $\Delta\Delta G$ data using practically-important metrics

 \rightarrow Both



Confusion matrix









Confusion matrix



Confusion matrix









Confusion matrix









Confusion matrix









Emergence of mutation scoring capabilities

Self-supervised pre-training





Pre-training is crucial for fine-tuning



600 1200 1800 0 Training step

Supervised $\Delta\Delta G$ fine-tuning

With pre-training

Without pre-training



Pre-training from PPIRef is crucial

Pre-training



DIPS/DIPS-Plus (deduplicated; 8K) DIPS/DIPS-Plus (40K) **PPIRef50K** (filtered, deduplicated) PPIRef300K (filtered) PPIRef800K (raw)

- Precision on negative $\Delta\Delta G$ (zero-shot) - Per-PPI Spearman on $\Delta\Delta G$ (zero-shot)





Other key ingredients

Pre-training



PPIformer

No 80%10%10% **BERT** masking Masking both chains With $N - C_{\alpha} - C$ frames With $C_{\alpha} - C_{\alpha} - C_{\alpha}$ frames No label smoothing $(\epsilon = 0)$ No class weights No class weights, No label smoothing

- Precision on negative $\Delta\Delta G$ (zero-shot) -•- Per-PPI Spearman on $\Delta\Delta G$ (zero-shot)



0.10 0.15 0.20 0.25 0.30 0.35



Comparison with the state of the art: 5 independent PPIs from SKEMPI

Category	Method	Spearman ↑	Pearson ↑	Precision \uparrow	Recall ↑	ROC AUC \uparrow	$MAE \downarrow$	$RMSE\downarrow$
Force field simulations	Flex dd G * ²²	0.54	0.57	0.63	0.62	0.84	1.60	2.00
Machine learning	MSA TRANSFORMER ²³	0.37	<u>0.45</u>	0.51	0.38	0.76	5.99	6.77
	ESM-IF ²⁴	0.32	0.31	0.36	0.28	0.69	1.84	2.11
	RDE-NET. ²¹	0.24	0.30	<u>0.54</u>	0.65	0.67	<u>1.70</u>	2.02
	PPIFORMER (OURS)	0.42	0.46	0.58	<u>0.61</u>	0.77	1.64	1.94

* A single prediction requires ~1 CPU hour (5 orders of magnitude slower than other methods).



Structural database of Kinetics and Energetics of Mutant Protein Interactions



Comparison with the state of the art: COVID

	k	nown fa	precision					
Method	тнз1₩↓	AH53F↓	NH57L↓	RH103M↓	LH104F↓	P@1↑	P@5%↑	P@10%↑
MSA TRANSFORMER	56.88	42.11	63.56	49.19	18.83	0.00	0.00	0.00
ROSETTA	10.73	76.72	93.93	13.56	6.88	0.00	0.00	2.04
FOLDX	5.67**	68.22	2.63	12.35	29.96	0.00	<u>4.00</u>	4.08
DDGPRED	2.02	14.17	24.49	4.05	6.48	0.00	8.00	6.12
END-TO-END	11.34	16.60	8.30	52.43	80.36	0.00	0.00	2.04
MIF-NET.	27.94	66.19	8.50	17.21	36.23	0.00	0.00	2.04
ESM-IF	49.39	17.61	17.00	51.42	48.58	0.00	0.00	0.00
RDE-NET.	1.62	2.02	20.65	61.54	5.47	0.00	8.00	6.12
PPIFORMER (OURS)	18.02	0.20	7.69	21.46	10.93	100	<u>4.00</u>	<u>4.08</u>

** Mutations that are in top 10% of predictions are in bold.





Comparison with the state of the art: stroke

known favorable mutations

	Mutations with $\geq 2 \times$ activity enhancement								
Method	KC74Q					KC74R	Activity enhancement		
Mediod	KC135R				KC130E	KC130T	Activity childheethent		
	KC130A↓	КС130Т↓	КС130Т↓	КС135А↓	KC135R↓	KC135R↓	P@1↑	P@5% ↑	P@10% ↑
MSA TRANSFORMER	52.50	32.50	55.00	40.0	70.00	78.75	100	50.00	37.50
ESM-IF	45.00	33.75	46.25	25.0	42.50	58.75	<u>0.00</u>	0.00	25.00
RDE-NET.	51.25	33.75	22.50	15.00	27.50	5.00	0.00	50.00	62.50
PPIFORMER (OURS)	66.25	15.00	2.50	52.50	33.75	1.25	100	75.00	87.50



precision

Microplasmin



LEARNING TO DESIGN PROTEIN–PROTEIN INTERAC-TIONS WITH ENHANCED GENERALIZATION

Anonymous authors Paper under double-blind review

Abstract

Discovering mutations enhancing protein–protein interactions (PPIs) is critical for advancing biomedical research and developing improved therapeutics. While machine learning approaches have substantially advanced the field, they often struggle to generalize beyond training data in practical scenarios. The contributions of this work are three-fold. First, we construct PPIRef, the largest and non-redundant dataset of 3D protein–protein interactions, enabling effective large-scale learning. Second, we leverage PPIRef to pre-train PPIformer, a new SE(3)-equivariant model generalizing across diverse protein-binder variants. We fine-tune PPIformer to predict effects of mutations on protein–protein interactions via a thermodynamically motivated adjustment of the pre-training loss function. Finally, we demonstrate the enhanced generalization of our new PPIformer approach by outperforming other state-of-the-art methods on the new non-leaking splits of the standard labeled PPI mutational data and independent case studies optimizing a human antibody against SARS-CoV-2 and increasing staphylokinase throm-bolytic activity.

PPIformer Github



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